

## Structure–Activity Relationships among DNA-Gyrase Inhibitors. Synthesis and Antimicrobial Evaluation of Chromones and Coumarins Related to Oxolinic Acid

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In the course of synthesizing 7-methoxychromone-3-carboxylic acid (*5a*) and 6,7-methylenedioxychromone-3-carboxylic acid (*5b*), as 1-oxa analogues of oxolinic acid (*1*), we investigated three different routes to chromone-3-carboxylates. Acid *5b* was most effectively achieved via a Kostanecki-Robinson reaction. Acid *5a* was obtained by NBS oxidation of the corresponding aldehyde *14a*, which was synthesized by an improved method utilizing a difluorodioxaborin intermediate *13*. Reaction between diethyl ethoxymethylenemalonate (*9a*) and 3-hydroxyanisole (*10*) has been studied under thermal and Lewis acid catalyzed conditions. Upon heating, small amounts of ethyl ester of chromone *5a* (*4a*), corresponding coumarin *6a*, and uncyclized addition product *9b* were found. No significant antimicrobial activity of the compounds tested was revealed.

Antibacterial agents of the oxolinic acid (*1*) and nalidixic acid (*2*) type possess a specific activity against DNA gyrase and have found a use in human medicine in the treatment of urinary tract

infections. Extensive structure activity studies in this class of compounds have been made.<sup>1</sup> Recently, we reported the synthesis and antimicrobial activity of a series of carbocyclic analogues (*3*).<sup>2</sup> This study clearly showed that replacement of the *N*-Et moiety of *1* by a Me–C–Me grouping diminishes activity against both bacteria and the semi-purified enzyme by several orders of magnitude. As a continuation of our studies on the importance of the nitrogen atom, we now report the synthesis and evaluation of some chromone (*4,5*) and coumarin (*6–8*) analogues.

It was reasoned that nitrogen, with its pronounced electron releasing character, was in an excellent position in these molecules (*1*) to contribute to electrophilicity driven reactions with the unsaturated  $\beta$ -ketocarboxylic acid system of the A ring (Fig. 1). On the other hand, the dimethylcarba analogues (*3*) could not do so. The 1-oxa analogues lie somewhere between these two extremes and therefore seemed worthy of study.

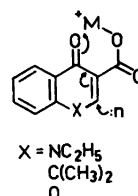
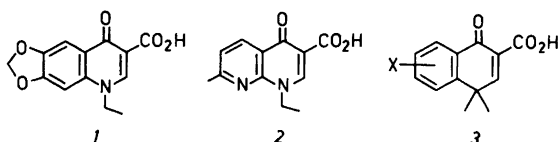


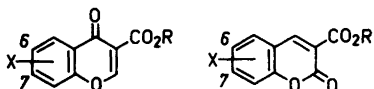
Fig. 1. Putative mechanism of action of DNA-gyrase inhibitors involves complexation of a metal cation possibly to facilitate Michael attack by bio-nucleophile.

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## CHEMISTRY

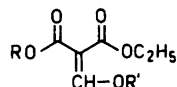
Compounds such as *1* and *2* can be made by reaction of the corresponding arylamine and diethyl ethoxymethylenemalonate (*9a*) at high temperatures.<sup>1</sup> A corresponding reaction between an activated phenol and *9a* would, in principle, be a simple way to the corresponding chromones (*e.g.*, *4*). However, such reactions have been studied and are claimed to lead to coumarins (*6*) in low yields.<sup>3</sup> This reaction was recently improved upon by Bissell through use of



R	X		
Et	7-OCH <sub>3</sub>	<i>4a</i>	<i>6a</i>
Et	6,7-OCH <sub>2</sub> O	<i>4b</i>	<i>6b</i>
H	7-OCH <sub>3</sub>	<i>5a</i>	<i>7a</i>
H	6,7-OCH <sub>2</sub> O	<i>5b</i>	<i>7b</i>
H	H	<i>5c</i>	<i>7c</i>

Lewis acid catalysis.<sup>4</sup> However, the inherent low yields in the above reactions and the possibility of formation of chromone via initial addition-elimination of the double bond in *9a* in analogy with reaction with anilines, prompted us to reinvestigate this process to see if the desired chromones could be formed in this way. We chose to study the reaction of 3-hydroxyanisole (*10*) and *9a* since the desired product (*4a*) has the methoxy group in the C-7 position known to be optimal for bioactivity based on structure-activity relationships established for quinolonecarboxylic acids.<sup>1</sup> Furthermore, a *meta*-substitution pattern is necessary for efficient ring closure to either a chromone or a coumarin system. We were indeed able to isolate a small amount (3 %) of chromone *4a* from the very complex reaction mixture resulting from thermal reaction of *9a* and *10*. However, to further complicate matters, the melting point (117–118 °C) of *4a* so obtained was identical to that reported for *6a* by Bissell resulting from a similar Lewis acid catalyzed reaction of *9a* and *10*.<sup>4</sup> Since it is well known to be difficult to differentiate between chromones and coumarins,<sup>5</sup> we felt it necessary to synthesize *6a* by an unambiguous Knoevenagel reaction and to repeat Bissell's reaction employing, however, a simplified work up. The coumarins *6a* obtained by the two routes had melting points of 134 °C in

accordance with earlier reports<sup>6</sup> and were identical to each other. We were, however, not able to detect any appreciable amounts of chromone *4a* by TLC or NMR in the crude product of the ZnCl<sub>2</sub> catalyzed Bissell reaction. In extending our work, we found that a similar thermal condensation of *9a* and *15* led to no isolable quantity of chromone *4b* but gave only small amounts of coumarin *6b* instead. When desired, the latter coumarin (*6b*) was preferably made by the Lewis acid catalyzed reaction which proceeded in this case in 42 % yield.



- 9a*. R=R'=Et  
*9b*. R=Et, R'=3-MeOC<sub>6</sub>H<sub>4</sub>  
*9c*. R=3-MeOC<sub>6</sub>H<sub>4</sub>, R'=Et

In the thermal reaction of *10* and *9a* we also isolated an uncyclized addition product in 10 % yield. <sup>1</sup>H NMR showed two sets of ethoxy signals, which clearly indicated that the structure was *9b* and not that of acylation product *9c*. The latter (*9c*) has the option of *Z/E* isomerism which should result in at least a small amount of two more sets of ethoxy signals. Furthermore, *9b* did not form either *4a* or *6a* upon refluxing for 2 h with one equivalent of ZnCl<sub>2</sub> in ethanol, but rather underwent ethanolysis to *10* and *9a*. Hydrogenation of *9b* in methanol gave diethyl methylmalonate, which confirms the structure to be *9b i. e.*, the first intermediate in the formation of chromone *4a*.

We also attempted a Lewis acid (ZnCl<sub>2</sub>) catalyzed condensation of *10* and *9a* in diphenyl ether at 200 °C, conditions similar to those successfully applied to the synthesis of 4-hydroxyquinoline-3-carboxylic esters.<sup>7</sup> The main compounds formed were coumarin *6a* and a symmetrical coumarinocoumarin *11* formed by *O*-acylation of *10* with *6a*, ring closure, and air-oxidation. The other alternatives, shown in Fig. 2, the chromonochromone and the unsymmetrical chromonocoumarin, can be ruled out primarily based upon the lack of a retro Diels-Alder fragment in the mass spectra (*vide infra*).<sup>5</sup> A trace amount of *11* dissolved in DMSO and the <sup>1</sup>H NMR of this solution showed a characteristic non-bonded interaction between the C-1 H and the C-12 H

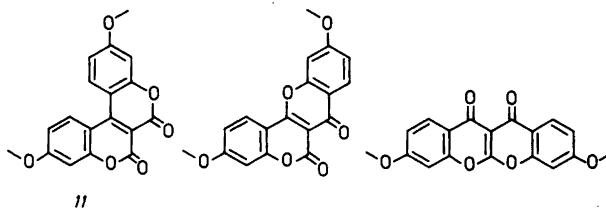
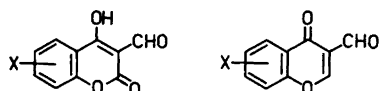


Fig. 2. Three possible [1]benzopyrano[1]benzopyran products from the reaction between diethyl ethoxymethylenemalonate and 3-methoxyphenol.

causing a down-field shift to 8.32 ppm similar to that observed in 3,4-benzophenanthrene.<sup>8,9</sup> The larger *ortho* coupling constant ( $J=9.5$  Hz) is also consistent with this type of annelation.<sup>9</sup> Furthermore, the unsubstituted chromonochromone and chromonocoumarin, made from bis(2-hydroxybenzoyl)methane,<sup>10</sup> have CO absorption frequencies in IR significantly lower than that of the coumarinocoumarin 11.

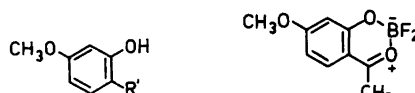
Basic hydrolysis of the coumarin esters gave 7a and 7b. However, when chromone-3-carboxylates (4) are subjected to basic conditions they rearrange, via initial Michael attack of hydroxide, to give 3-formyl-4-hydroxycoumarins (8).<sup>11</sup> We hydrolyzed 4a with hydrochloric acid to the known acid 5a and with base to 8a because the



X=7-OCH<sub>3</sub>      8a      14a  
 X=6,7-OCH<sub>2</sub>O      8b      14b

enolic system in 8a is acidic<sup>11</sup> enough to render it an interesting structure, related to 7a, to investigate biologically.

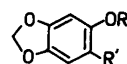
An alternative route to chromone-3-carboxylic acids *via* the corresponding aldehyde has been devised by Nohara *et al.*<sup>12</sup> However, the Vilsmeier-Haack type reaction with acetophenone 12 and DMF-POCl<sub>3</sub> is reported to give the desired aldehyde 14a in very low yield (6%),<sup>12</sup> which proved to be the case in our hands also. Similarly, we were only able to obtain aldehyde 14b in low yield (7%) from 16. This is consistent with the reported low yield reaction of 4,5-dimethoxy-2-hydroxyacetophenone (4%).<sup>12</sup> To overcome the problems of using this type of activated acetophenone as starting materials, we next investigated the method reported by Reynolds *et al.* for the



10. R'=H  
 12. R'=COCH<sub>3</sub>

preparation of 3-formylbenzo[*h*]-chrom-4-one from 2,2-difluoro-4-methylnaphtho[2,1-*e*]-1,3,2-dioxaborin.<sup>13</sup>

Thus 12 was reacted with boron trifluoride etherate in acetic anhydride to give the difluorodioxaborin 13 in 85% yield. Treatment of 13 with Vilsmeier reagent in DMF yielded aldehyde 14a in 94% yield. Oxidation of 14a was then accomplished by *N*-bromosuccinimide followed by quenching with water in accordance with Machida *et al.*<sup>14</sup> to give 5a. We were not, however, able to oxidize the methylenedioxy



	R	R'
15	H	H
16	H	COCH <sub>3</sub>
17	CH <sub>2</sub> Ph	COCH <sub>3</sub>
18	CH <sub>2</sub> Ph	COCH <sub>2</sub> CO <sub>2</sub> Et
19	H	COCH <sub>2</sub> CO <sub>2</sub> Et

aldehyde 14b to produce useful quantities of the acid despite exploration of various conditions.

Therefore, an alternative route from the same acetophenone 16, following the outline of Okumura's synthesis of 5c,<sup>15</sup> was employed. Phenol 16 was benzylated to 17 and reacted with diethyl carbonate and sodium hydride to give 18 in good yield. Following deprotection to 19, Kostanecki-Robinson type reaction with acetic formic anhydride/sodium formate in THF gave 4b in high yield. Acidic and basic hydrolysis of 4b as

Table 1. Structure elucidation by  $^1\text{H}$  NMR and mass spectrometry.

No.	$\delta(\text{ppm})$		Prominent MS fragments; $m/z$ (rel. intensity)
	C-5 H	C-2 H, C-4 H, or CHO	
6a	7.51	8.50	248(M, 53 %), 148(M-CO <sub>2</sub> Et+H-CO, 25 %)
4a	8.16	8.57	248(M, 10 %), 151(retro DA+H, 24 %), 150(4 %)
6b	6.91	8.42	262(M, 100 %), 162(M-CO <sub>2</sub> Et+H-CO, 68 %)
4b	7.58	8.55	262(M, 17 %), 165(25 %), 164(retro DA, 14 %)
7a	7.50	8.86	220(M, 85 %), 176(M-CO <sub>2</sub> , 100 %), 148(M-CO <sub>2</sub> -CO, 68 %)
5a	8.23	8.92	220(M, 3 %), 176(M-CO <sub>2</sub> , 100 %), 150(retro DA, 11 %)
8a	7.94	9.97	220(M, 54 %), 192(M-CO, 100 %), 150(retro DA, 98 %)
7b	7.10	8.80	234(M, 46 %), 190(M-CO <sub>2</sub> , 32 %), 162(M-CO <sub>2</sub> -CO, 100 %)
5b	7.58	8.89	234(M, 13 %), 190(M-CO <sub>2</sub> , 100 %), 164(retro DA, 31 %)
8b	7.31	9.97	234(M, 26 %), 206(M-CO, 27 %), 164(retro DA, 100 %)
14b	7.59	8.45	218(M, 8 %), 190(M-CO, 100 %), 164(retro DA, 40 %)
		10.38	

described above gave 5b and 8b, respectively.

The problems often encountered in distinguishing chromones and coumarins<sup>5</sup> lead us to comment on the structural proofs of 4-8 and 14. Table 1 outlines some of the important features found in their  $^1\text{H}$  NMR and mass spectra. The C-5 H has a pronounced downfield shift in the chromones and in the 4-hydroxycoumarins (8) an intermediate effect is observed. In the mass spectra, the retro Diels-Alder fragmentation is of great diagnostic value for the chromones whereas the ring contraction by elimination of carbon monoxide is more characteristic for the coumarins. In the case of 8, a marked retro Diels-Alder fragmentation is observed due to the tautomerism with the 2-hydroxy-3-formyl-chromone.

## BIOLOGY

The *in vitro* antimicrobial assays were carried out by the standard agardilution/streak technique with a highest test level of 250  $\mu\text{g}/\text{ml}$ .<sup>16</sup> The compounds 4b, 5a, 5b, 6a, 6b, 7a, 7b, 7c, 8a and 8b were tested on inhibition of *Staphylococcus aureus* (ATCC 13709), *Escherichia coli* (ATCC 9637), *Salmonella gallinarum* (ATCC 9184), *Klebsiella pneumoniae* AD (ATCC 10031), *Mycobacterium smegmatis* (ATCC 607) and *Candida albicans* (ATCC 10231). However, no appreciable inhibition was observed with the exception of 8a, which inhibited *Staph. aureus* and *M. smegmatis* at the level of 12.5  $\mu\text{g}/\text{ml}$ .

These results further underline the unique properties of the *N*-Et moiety in antimicrobial agents such as 1 and 2.

## EXPERIMENTAL

Melting points were determined on a Thomas-Hoover apparatus and are uncorrected. Infrared spectra were recorded on a Beckman 33 spectrophotometer. Mass spectra were conducted on a Hitachi Perkin-Elmer RMS-4 mass spectrometer (EI, 70 eV).  $^1\text{H}$  NMR spectra were recorded on a Varian FT 80-A spectrometer with tetramethylsilane as internal standard. Microanalyses were performed on a Hewlett-Packard 185B at the University of Kansas and the results are within  $\pm 0.4$  % of the theoretical values.

*Ethyl 6,7-methylenedioxy coumarin-3-carboxylate* (6b). A solution of sesamol (15, 2.76 g, 0.02 mol), diethyl ethoxymethylenemalonate (9a, 6.48 g, 0.03 mol) and dry  $\text{ZnCl}_2$  (3.4 g, 0.025 mol) in 10 ml EtOH was refluxed for 23 h. After addition of aqueous HCl, the mixture was extracted several times with  $\text{CH}_2\text{Cl}_2$ . The organic phase was dried ( $\text{Na}_2\text{SO}_4$ ) and evaporated to give 5.8 g of a residue. Repeated recrystallization from EtOH/EtOAc afforded 2.20 g (42 %) pure coumarin 6b, m.p. 192.5-194 °C.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ):  $\delta$  1.39 (t), 4.39 (q), 6.12 (s,  $\text{OCH}_2\text{O}$ ), 6.80 (s), 6.91 (s), 8.42 (s, 4-H); MS (EI):  $m/z$  (rel. int.) 262 (M, 100), 234 (M-CO, 8), 217 (M-C<sub>2</sub>H<sub>5</sub>O, 77), 206 (30), 205 (23), 190 (M-CO<sub>2</sub>C<sub>2</sub>H<sub>5</sub>+H, 84), 162 (190-CO, 68), 133

(52); Anal.  $C_{13}H_{10}O_6$ : C, H.

*Ethyl 7-methoxycoumarin-3-carboxylate (6a)* was prepared in analogy with the above procedure from 3-methoxyphenol (*10*) and *9a* (1:1 ratio) in 9% yield, m.p. 133–134 °C (lit.<sup>4</sup> 25%, m.p. 117 °C). It was also prepared according to Rangaswami *et al.*<sup>6a</sup> from 2-hydroxy-4-methoxybenzaldehyde and diethylmalonate in 64% yield, m.p. 134–135 °C (lit.<sup>6a,6b</sup> 134 °C). <sup>1</sup>H NMR ( $CDCl_3$ ):  $\delta$  1.40 (t), 3.90 (s), 4.40 (q), 6.82 (m, 8-H), 6.87 (m, 6-H), 7.51 (d,  $J=8.1$  Hz, 5-H), 8.50 (s, 4-H); MS (EI):  $m/z$  (rel. int.) 248 (M, 53), 220 (M–CO, 4), 203 (M– $C_2H_5O$ , 100), 176 (M– $CO_2C_2H_5+H$ , 86), 148 (176–CO, 25).

*6,7-Methylenedioxcoumarin-3-carboxylic acid (7b)*. The ester *6b* (262 mg, 1 mmol) was stirred in 5 ml 2 M NaOH and 10 ml EtOH for 3.5 h at room temperature. The ethanol was evaporated and the yellow suspension was acidified with conc. HCl and stirred for 4 h. The precipitate was filtered and washed with water. Recrystallization from EtOH/acetone afforded 205 mg (88%) acid, m.p. 275–276 °C. <sup>1</sup>H NMR ( $CDCl_3/CD_3CN$ ):  $\delta$  6.18 (s,  $OCH_2O$ ), 6.95 (s, 8-H), 7.10 (s, 5-H), 8.80 (s, 4-H); MS (EI):  $m/z$  (rel. int.) 234 (M, 46), 206 (M–CO, 10), 205 (23), 190 (M– $CO_2$ , 32), 162 (M– $CO_2-CO$ , 100), 161 (86), 134 (11), 133 (24), 132 (16), Anal.  $C_{11}H_6O_6$ : C, H.

*7-Methoxycoumarin-3-carboxylic acid (7a)* was prepared by alkaline hydrolysis of the ethyl ester *6a*, m.p. 192–194 °C (lit.<sup>6a,6b</sup> 195 °C); <sup>1</sup>H NMR ( $CDCl_3$ ):  $\delta$  3.96 (s), 6.9 and 7.0 (overlapping d and dd,  $J_{6,8}=2.4$  Hz, 6-H and 8-H), 7.50 (d,  $J=8.5$  Hz, 5-H), 8.86 (s, 4-H), 12.20 (br s, OH); MS (EI):  $m/z$  (rel. int.) 220 (M, 85), 203 (17), 192 (M–CO, 8), 177 (49), 176 (M– $CO_2$ , 100), 149 (15), 148 (M– $CO_2-CO$ , 68), 134 (38), 133 (99).

*Coumarin-3-carboxylic acid (7c)* was prepared by alkaline hydrolysis of the methyl ester, m.p. 189–191 °C (lit.<sup>17</sup> 191–192 °C).

*Ethyl 7-methoxychromone-3-carboxylate (4a)*. A. A mixture of 3-methoxyphenol (*10*, 1.24 g, 0.01 mol) and diethyl ethoxymethylenemalonate (*9a*, 6.49 g, 0.03 mol) was heated at 170 °C for 2 days. TLC showed a complex reaction mixture containing at least 6 components including a small amount of coumarin (*6a*). Most of the excess of malonate was distilled off in vacuo and the residue separated by flash chromatography on  $SiO_2$  with 3 l (1:3) and 1 l (1:1) EtOAc–hexane to give *4a* and *9b*. The chromone *4a* was isolated in 2.9% yield, m.p. 117–118 °C. <sup>1</sup>H NMR ( $CDCl_3$ ):  $\delta$  1.38 (t), 3.90 (s), 4.38 (q), 6.86 (d,  $J=2.3$  Hz, 8-H), 6.99 (dd,  $J=8.9$  and 2.3 Hz, 6-H), 8.16 (d,  $J=8.9$  Hz, 5-H), 8.57 (s, 2-H); MS (EI):  $m/z$  (rel. int.)

248 (M, 10), 203 (M– $C_2H_5O$ , 27), 176 (M– $CO_2C_2H_5+H$ , 100), 151 (retro Diels–Alder +H, 24), 150 (4), 148 (1.6): Peak match: Found  $m/z$  248.06666. Calc. for  $C_{13}H_{12}O_5$  248.06839.

*Open adduct 9b* was isolated in 10% yield. <sup>1</sup>H NMR ( $CDCl_3$ ):  $\delta$  1.30 and 1.34 (two t), 3.79 (s), 4.26 and 4.35 (two q), 6.6 (m), 7.27 (ca. dd,  $J=8.4$  Hz, 5-H), 7.87 (s). MS (EI):  $m/z$  (rel. int.) 294 (M, 79), 249 (M– $C_2H_5O$ , 64), 248 (48), 220 (34), 203 (55), 192 (15), 177 (14), 176 (19), 175 (27), 149 (13), 148 (27), 135 (100); Anal.  $C_{15}H_{18}O_6$ : C, H.

B. A mixture of *10* (2.48 g, 20 mmol) and *9a* (8.64 g, 40 mmol) was heated for 12 h at 205 °C. Work-up as above gave 2.2% *4a* and 6.8% *9b*.

*Hydrogenation of 9b*. The adduct *9b* (100 mg, 0.4 mmol) was hydrogenated over 5% Pd/C (0.05 g) in 50 ml MeOH at 0.28 MPa for 1.5 h.  $CH_2Cl_2$  was added and the solution was filtered and evaporated. The residue was dissolved in  $CH_2Cl_2$  and filtered through  $SiO_2$  plug and eluted with  $CH_2Cl_2$  to remove phenol *10*. Evaporation gave 47 mg (68%) diethyl methylmalonate. <sup>1</sup>H NMR ( $CDCl_3$ ):  $\delta$  1.27 (t), 1.40 (d), 3.43 (q), 4.21 (q), MS (EI):  $m/z$  (rel. int.) 174 (M, 6), 129 (M– $OC_2H_5$ , 100).

*6H,7H-3,10-Dimethoxy[1]benzopyrano[3,4-c]-[1]benzopyran-6,7-dione (11)*. A mixture of 3-methoxyphenol (*10*, 1.24 g, 10 mmol), diethyl ethoxymethylenemalonate (*9a*, 3.24 g, 15 mmol),  $ZnCl_2$  (100 mg, 0.7 mmol) and 6 g diphenyl ether was heated to 200 °C for 1 h. After the first 10 min a sudden precipitate developed. After cooling, hexane was added and the solvent decanted. The precipitate proved largely insoluble in most solvents (EtOAc,  $CH_2Cl_2$ ,  $Et_2O$ ). The precipitate was triturated with hot EtOH, filtered and washed with EtOH and finally  $Et_2O$  to give 0.30 g (0.9 mmol) weakly yellow precipitate, m.p. 317–319 °C (dec.). <sup>1</sup>H NMR ( $DMSO-d_6$ ):  $\delta$  3.95 (s), 7.1 (m, 2- and 11-H), 7.14 (br s, 4- and 9-H), 8.32 (d,  $J=9.5$  Hz, 1- and 12-H); IR (KBr): 1765, 1605  $cm^{-1}$ ; MS (EI):  $m/z$  (rel. int.) 324 (M, 100), 323 (43), 296 (M–CO, 27), 281 (M–CO– $CH_3$ , 17), 268 (M–CO–CO, 59), 253 (M–CO–CO– $CH_3$ , 32), 238 (11), 237 (52). Peak match:  $m/z$  Found 324.0641. Calc. for  $C_{18}H_{12}O_6$  324.0633.

*2,2-Difluoro-7-methoxy-4-methylbenzo[e]-1,3,2-dioxaborin (13)*. 2-Hydroxy-4-methoxyacetophenone (*12*, 100 g, 0.6 mol) and boron trifluoride etherate (150 ml, 1.2 mol) were added to 230 ml acetic anhydride. The solution was heated at 100 °C for 1 h. The precipitate was collected and washed with  $Et_2O$  three times to give 109.7 g (85%) of *13*, m.p. 170 °C; <sup>1</sup>H NMR ( $CDCl_3$ ):  $\delta$  2.70 (s,  $CH_3$ ), 3.93 (s,  $OCH_3$ ), 6.53

(d,  $J=2.2$  Hz, 8-H), 6.60 (dd,  $J=2.2$  and 8.8 Hz, 6-H), 7.70 (d,  $J=8.8$  Hz, 5-H). Anal.  $C_9H_9BF_2O_3$ : C, H.

**7-Methoxychromone-3-carboxaldehyde (14a).** To 50 ml DMF at 0 °C was added  $POCl_3$  (6.5 ml, 70 mmol) followed by difluorodioxaborin **13** (5 g, 23 mmol). The solution was heated at 100 °C for 1 h, cooled and poured into 600 ml ice water. After storing in refrigerator the precipitate was collected, washed with cold water and dried. Recrystallization from DMF yielded 4.5 g (94 %) of **14a**, m.p. 188 °C (lit.<sup>11</sup> 188–190 °C);  $^1H$  NMR ( $CDCl_3$ ): 3.95 (s), 7.01 (d,  $J=2.2$  Hz, 8-H), 7.06 (dd,  $J=2.2$  and 8.8 Hz, 6-H), 8.25 (d,  $J=8.8$  Hz, 5-H), 8.47 (s, 2-H), 10.40 (s, CHO). Anal.  $C_{11}H_8O_4$ : C, H.

**6,7-Methylenedioxychromone-3-carboxaldehyde (14b)** was prepared in analogy with Nohara *et al.*<sup>12</sup> A solution of 2-hydroxy-4,5-methylenedioxyacetophenone<sup>18</sup> (**16**, 541 mg, 3 mmol) in 15 ml dry DMF was cooled in an ice bath and freshly distilled  $POCl_3$  (0.92 ml, 10 mmol) was injected under Ar over 1 h. The mixture was stirred at 0 °C for 2 h and at room temperature over night and then poured into ice water. The aqueous phase was extracted with EtOAc, dried ( $MgSO_4$ ) and evaporated. Flash chromatography on  $SiO_2$  with 25 % EtOAc in hexane afforded 48 mg (7 %) aldehyde, m.p. 227–230 °C;  $^1H$  NMR ( $CDCl_3$ ):  $\delta$  6.15 (s), 6.92 (s, 8-H), 7.59 (s, 5-H), 8.45 (s, 2-H), 10.38 (s, CHO); MS (EI):  $m/z$  (rel. int.) 218 (M, 8), 190 (M–CO, 100), 189 (51), 164 (retro-Diels-Alder, 40); Peak match: Found  $m/z$  218.02034. Calc. for  $C_{11}H_6O_5$  218.02147.

**2-Benzyloxy-4,5-methylenedioxyacetophenone (17).** A mixture of 2-hydroxy-4,5-methylenedioxyacetophenone<sup>18</sup> (**16**, 895 mg, 5 mmol), benzyl bromide (1 ml, 8.4 mmol) and  $K_2CO_3$  (1.32 g, 9.5 mmol) in 30 ml dry acetonitrile was refluxed overnight. TLC ( $FeCl_3$  spray) indicated some unreacted phenol so more benzyl bromide (0.35 ml) and  $K_2CO_3$  (0.45 g) were added and reflux was continued another night. After evaporation of the solvent, water was added and the aqueous phase extracted with  $Et_2O$ . The ethereal layer was washed with aqueous NaOH, dried ( $MgSO_4$ ) and evaporated (1.64 g). Flash chromatography on  $SiO_2$  with 25 % EtOAc in hexane gave 693 mg (52 %) crystalline product. Recrystallization of an analytical sample from EtOAc–hexane gave **17**, m.p. 117–118 °C.  $^1H$  NMR ( $CDCl_3$ ):  $\delta$  2.53 (s), 5.10 (s,  $CH_2Ph$ ), 5.96 (s,  $OCH_2O$ ), 6.57 (s, 3-H), 7.33 (s, 6-H), 7.39 (s, Ph); MS (EI):  $m/z$  (rel. int.) 270 (M, 6.6), 228 (4.4), 227 (2.6), 165 (2.7), 91 (100); Anal.  $C_{16}H_{14}O_4$ : C, H.

**Ethyl (2-benzyloxy-4,5-methylenedioxyben-**

**zoyl)acetate (18):** A mixture of **17** (615 mg, 2.27 mmol), diethyl carbonate (0.51 ml, 6 mmol) and NaH (50 % oil dispersion, 0.38 g, 8 mmol) in 25 ml THF was refluxed overnight. TLC showed complete conversion. The mixture was cooled and 1 ml HOAc and  $C_6H_6$  were added. After evaporation the residue was taken up in  $Et_2O$ , washed with water and brine. Drying ( $Na_2SO_4$ ) and evaporation gave a residue which was recrystallized from  $Et_2O$  to give 620 mg (80 %) pure product, m.p. 93–94.5 °C.  $^1H$  NMR ( $CDCl_3$ ):  $\delta$  1.26 (t), 3.91 (s,  $COCH_2CO$ ), 4.07 (q), 5.11 (s,  $CH_2Ph$ ), 5.97 (s,  $OCH_2O$ ), 6.53 (s, 3-H), 7.38 (s, Ph), 7.39 (s, 6-H); MS (EI):  $m/z$  (rel. int.) 342 (M, 1.9), 296 (M– $C_2H_5OH$ , 4.8), 165 (4.7), 164 (6.3), 91 (100); Anal.  $C_{19}H_{18}O_6$ : C, H.

**Ethyl (2-hydroxy-4,5-methylenedioxybenzoyl)acetate (19):** A solution of **18** (439 mg, 1.28 mmol) in 250 ml EtOH was hydrogenated over 5 % Pd/C (0.3 g) for 25 min at 0.25 MPa at room temperature. Filtration and evaporation gave a residue which was dissolved in  $Et_2O$  and filtered through a small  $SiO_2$  plug. Evaporation, addition of  $CH_2Cl_2$  and evaporation gave 323 mg (100 %) of an oil which crystallized, m.p. 61–63 °C.  $^1H$  NMR ( $CDCl_3$ ):  $\delta$  1.27 (t), 3.86 (s,  $COCH_2CO$ ), 4.22 (q), 5.98 (s,  $OCH_2O$ ), 6.43 (s, 3-H), 6.98 (6-H), 12.64 (s, OH); MS (EI):  $m/z$  (rel. int.) 252 (M, 18), 206 (M– $C_2H_5OH$ , 48), 165 (55), 164 (100); Anal.  $C_{12}H_{12}O_6$ : C, H.

**Ethyl 6,7-methylenedioxychromone-3-carboxylate (4b).** A mixture of **19** (194 mg, 0.77 mmol), sodium formate (68 mg, 1 mmol) and acetic formic anhydride (0.25 ml, ca. 1.75 mmol) in 1.5 ml dry THF was stirred at room temperature under Ar. TLC showed incomplete conversion after 4 h and another 0.25 ml acetic formic anhydride was injected, and the stirring continued overnight. The mixture was poured into ice water and extracted twice with  $CH_2Cl_2$ . The organic layer was washed with water and brine, dried ( $Na_2SO_4$ ), and evaporated to give 190 mg (94 %) of a white solid. Recrystallization from  $Et_2O$  gave 151 mg (75 %), m.p. 177–179 °C.  $^1H$  NMR ( $CDCl_3$ ):  $\delta$  1.39 (t), 4.39 (q), 6.11 (s,  $OCH_2O$ ), 6.87 (s, 8-H), 7.58 (s, 5-H), 8.55 (s, 2-H); MS (EI):  $m/z$  (rel. int.) 262 (M, 17), 217 (M– $C_2H_5O$ , 20), 190 (M– $CO_2C_2H_5+H$ , 100), 189 (24), 165 (25), 164 (retro Diels-Alder, 14), 133 (4); Anal.  $C_{13}H_{10}O_6$ : C, H.

**7-Methoxychromone-3-carboxylic acid (5a).** A. The ester **4a** (15 mg, 0.060 mmol) was refluxed in 4 ml 50 % aqueous HCl for 15 minutes and then diluted with some water and filtered. The filter cake was washed with  $H_2O$ , EtOH and finally  $Et_2O$ . Drying gave 8 mg (61 %) pure acid **5a**, m.p. 205–206.5 °C, with same m.p. after recryst-

tallization from acetone-hexane (lit.<sup>14</sup> 212–215 °C). By extraction of the aqueous filtrate with CH<sub>2</sub>Cl<sub>2</sub> another 4.8 mg (36 %) acid was obtained.

B. To a suspension of aldehyde *14a* (5 g, 25 mmol) in 150 ml alcohol free CHCl<sub>3</sub> was added *N*-bromosuccinimide (9 g, 50 mmol) and the mixture was illuminated by a flood lamp for 2 h with gentle refluxing. After cooling the solution was extracted with aqueous NaHCO<sub>3</sub>. The aqueous extract was acidified and extracted twice with CH<sub>2</sub>Cl<sub>2</sub>. After drying and evaporation the residue was recrystallized from acetonitrile to give 1.67 g (30 %) acid *5a*, m.p. 198–200 °C.

The acids prepared by the two routes were identical. <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ 3.96 (s), 6.99 (d, *J*=2.2 Hz, 8-H), 7.13 (dd, *J*=2.2 and 8.9 Hz, 6-H), 8.23 (d, *J*=8.9 Hz, 5-H), 8.92 (s, 2-H), 13.64 (s, OH); MS (EI) *m/z* (rel. int.): 220 (M, 3), 176 (M-CO<sub>2</sub>, 100), 151 (7), 150 (retro Diels-Alder, 11), 134 (25). Peak match: Found *m/z* 220.03682. Calc. for C<sub>11</sub>H<sub>8</sub>O<sub>5</sub> 220.03711.

*6,7-Methylenedioxychromone-3-carboxylic acid (5b)*: The ester *4b* (26 mg, 0.10 mmol) was refluxed in 2 ml 50 % aqueous HCl for 5 min and then diluted with ice water and filtered. The precipitate was washed with water and finally Et<sub>2</sub>O. Drying gave 16 mg (68 %) pure acid, m.p. 232–233 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ 6.20 (s, OCH<sub>2</sub>O), 7.00 (s, 8-H), 7.58 (s, 5-H), 8.89 (s, 2-H), 13.69 (s, OH); MS (EI) *m/z* (rel. int.) 234 (M, 13), 190 (M-CO<sub>2</sub>, 100), 189 (52), 164 (retro Diels-Alder, 31), 165 (6); Peak match: Found *m/z* 234.01749. Calc. for C<sub>11</sub>H<sub>6</sub>O<sub>6</sub>: 234.01638; Anal. C<sub>11</sub>H<sub>6</sub>O<sub>6</sub>: C, H.

*4-Hydroxy-7-methoxycoumarin-3-carboxaldehyde (8a)*: A mixture of *4a* (64 mg, 0.26 mmol), 2 ml 2 M NaOH and 10 ml EtOH was stirred at room temperature for 2 h. The ethanol was evaporated and aqueous NaHCO<sub>3</sub> was added and the aqueous phase was washed with Et<sub>2</sub>O twice. After acidification with H<sub>2</sub>SO<sub>4</sub> the aqueous phase was extracted with CH<sub>2</sub>Cl<sub>2</sub>. Drying (Na<sub>2</sub>SO<sub>4</sub>) and evaporation gave 53 mg (93 %) orange *8a*. Recrystallization from EtOAc gave an analytical sample, m.p. 215–217 °C (dec.). <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ 3.92 (s), 6.78 (d, *J*=2.3 Hz, 8-H), 6.91 (dd, *J*=2.3 and 8.8 Hz, 6-H), 7.94 (d, *J*=8.8 Hz, 5-H), 9.97 (s, CHO), 14.5 (br, OH); IR (CHCl<sub>3</sub>): 1720 (CHO), 1615 (pyrone CO); MS (EI) *m/z* (rel. int.) 220 (M, 54), 192 (M-CO, 100), 151 (80), 150 (retro Diels-Alder, 98), 122 (38), 107 (26); Peak match: Found *m/z* 220.03651. Calc. for C<sub>11</sub>H<sub>8</sub>O<sub>5</sub> 220.03711.

*4-Hydroxy-6,7-methylenedioxy coumarin-3-carboxaldehyde (8b)*. A mixture of *4b* (47 mg, 0.18 mmol), 5 ml 2 M NaOH and 5 ml EtOH was

stirred at room temperature for 45 min. The ethanol was evaporated and the residue was acidified with aqueous HCl and extracted twice with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was washed with brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and evaporated to give 37 mg (89 %). Trituration with hot EtOH gave 32 mg (76 %) pink product, m.p. 270 °C (dec.). <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ 6.14 (s, OCH<sub>2</sub>O), 6.79 (s, 8-H), 7.31 (s, 5-H), 9.97 (s, CHO); MS (EI) *m/z* (rel. int.) 234 (M, 26), 206 (M-CO, 27), 164 (retro Diels-Alder, 100), 136 (164-CO, 18); Peak match: Found *m/z* 234.01692. Calc. for C<sub>11</sub>H<sub>6</sub>O<sub>6</sub> 234.01638; Anal. C<sub>11</sub>H<sub>6</sub>O<sub>6</sub>: C, H.

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## REFERENCES

- Albrecht, R. *Prog. Drug Res.* 21 (1977) 9.
- Högberg, T., Khanna, I., Drake, S. D., Mitscher, L. A. and Shen, L. L. *J. Med. Chem.* 27 (1984) 306.
- Gerphagnon, M. C., Molho, D. and Mentzer, C. C. *R. Acad. Sci. Paris* 246 (1958) 1701.
- Bissell, E. R. *Synthesis* (1982) 846.
- Ellis, G. P. In Ellis, G. P., Ed., *Chromenes, Chromanones and Chromones*, Wiley, New York 1977, Chapters VIII and XX.
- a. Rangaswami, S., Seshadri, T. R. and Venkateswarly, V. *Proc. Indian Acad. Sci. Sect. A* 13 (1941) 316; b. Baker, W. and Collis, C. B. *J. Chem. Soc.* (1949) S12.
- Egri, J., Halmos, J. and Rakoczi, J. *Acta Chim. Acad. Sci. Hung.*, Tomus 73 (1972) 469.
- Cheney, B. V. *J. Am. Chem. Soc.* 90 (1968) 5386.
- Haigh, C. W. and Mallion, R. B. *Mol. Phys.* 22 (1971) 945 and 955.
- Eiden, F. and Schweiger, H.-D. *Synthesis* (1974) 511.
- Klutchko, S., Shavel, J., Jr. and von Strandtmann, M. *J. Org. Chem.* 39 (1974) 2436.
- Nohara, A., Umetani, T. and Sanno, Y. *Tetrahedron* 30 (1974) 3553.
- Reynolds, G. A. and VanAllan, J. A. *J. Heterocyclic Chem.* 6 (1969) 375.
- Machida, Y., Nomoto, S., Negi, S., Ikuta, H. and Saito, I. *Synth. Commun.* 10 (1980) 889.

15. Okumura, K., Kondo, K., Oine, T. and Inoue, I. *Chem. Pharm. Bull.* 22 (1974) 331.
16. Mitscher, L. A., Leu, R., Bathala, M. S., Wu, W.-N., Beal, J. L. and White, R. *Lloydia* 35 (1972) 157.
17. Clinton, R. O. and Laskowski, S. C. *J. Am. Chem. Soc.* 71 (1949) 3602.
18. Fukui, K. and Nakayama, M. *Bull. Chem. Soc. Jpn.* 37 (1964) 300.

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